

Protein X-ray crystallography and glycobiology

Dr Annabelle Varrot

« Structural Glycosciences » Summer School
IBS, Grenoble 7th June 2023

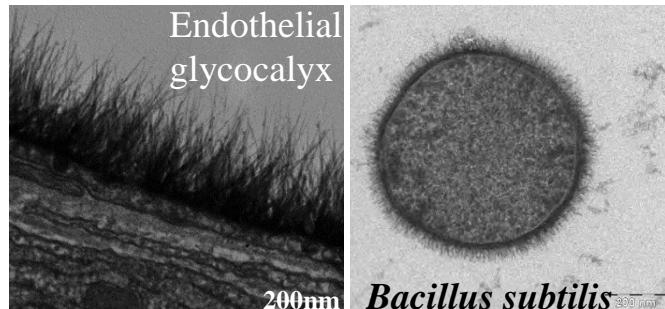




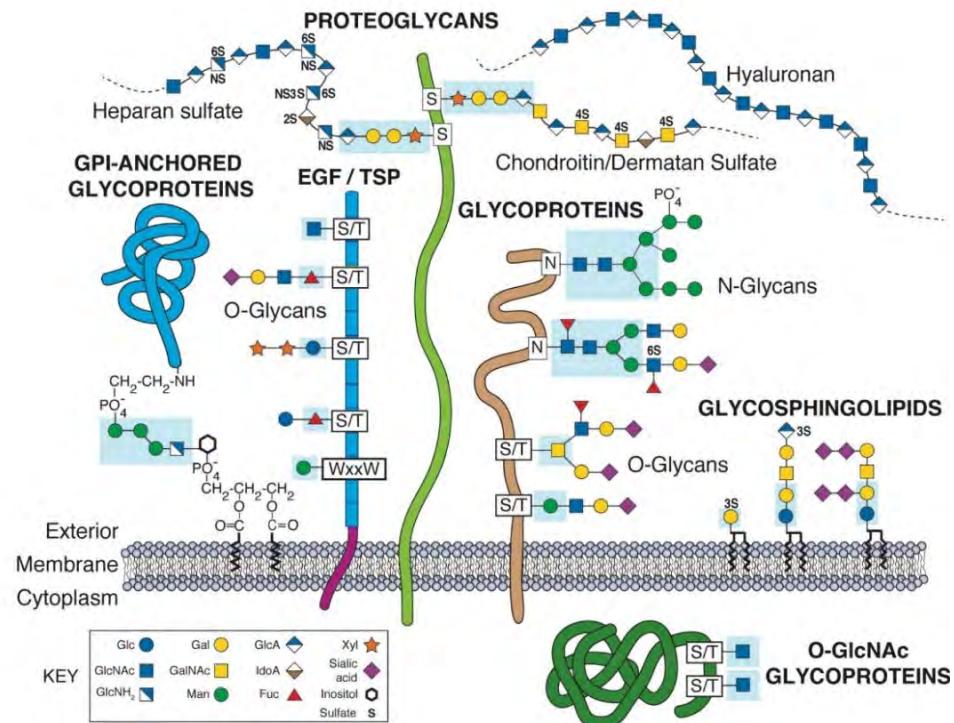
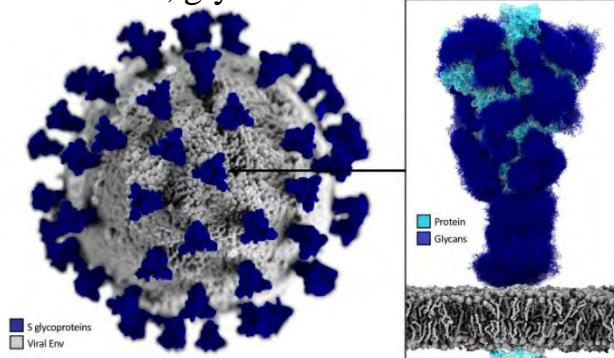
"Sweet side" of cells



- Sugars: 3rd alphabet of life: Glycome → glycocode
- Every cell is covered by **glycoconjugates**



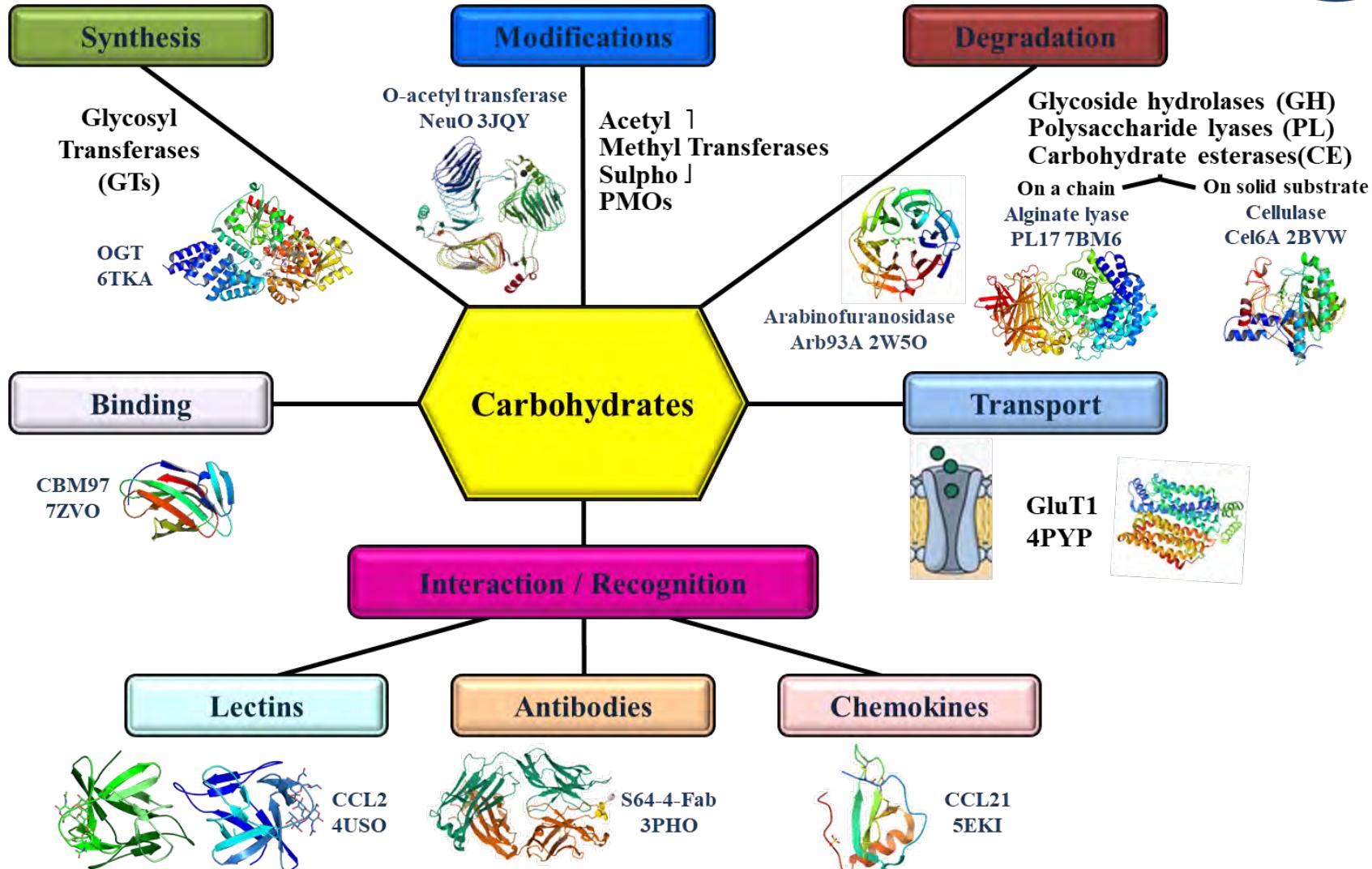
SARS-CoV-2, glycan shield



- Essentials in cell identity, fate, recognition & signaling

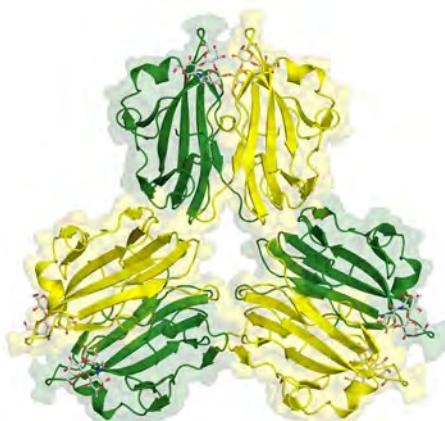


Protein-carbohydrate interactions



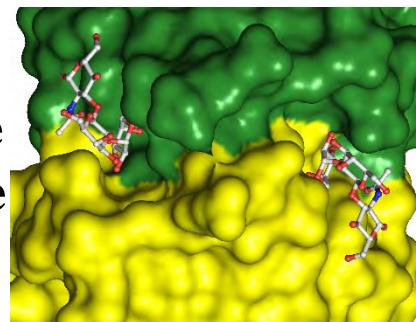


Which info by X-ray crystallography?

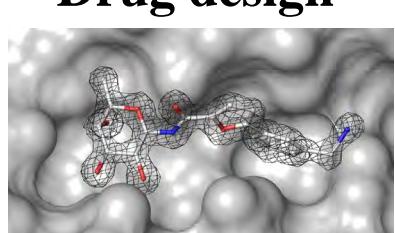
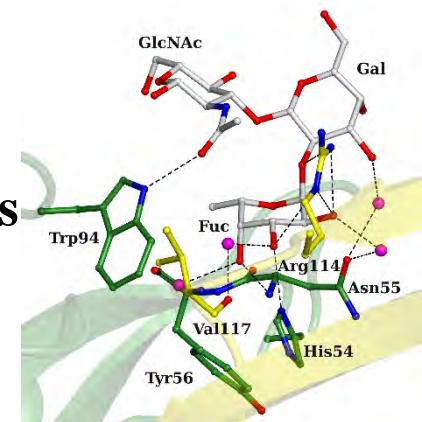


Fold/assembly

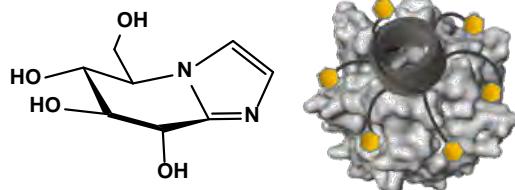
Binding site architecture



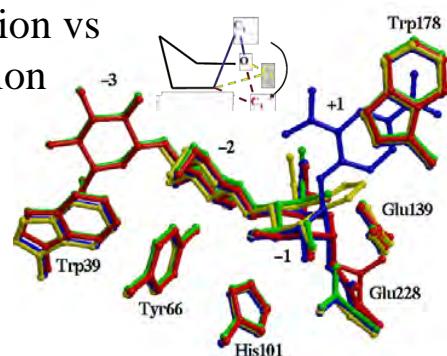
Interactions



Drug design



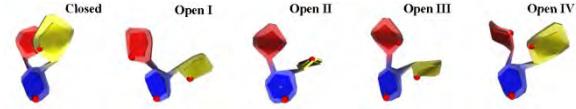
Retention vs
Inversion



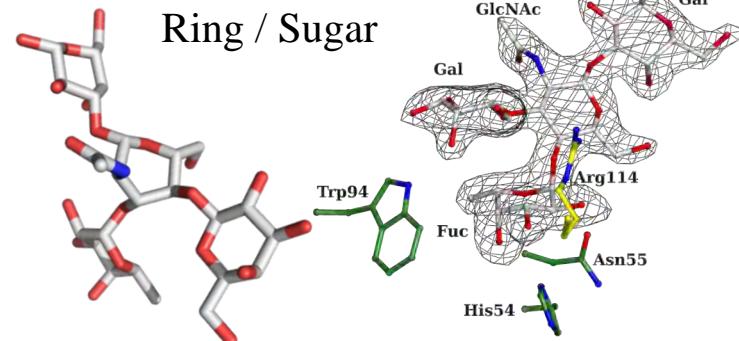
Structure

Enzyme mechanism

Conformation



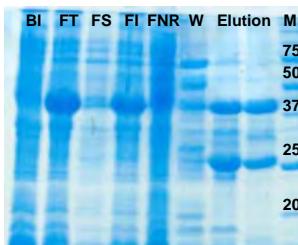
Ring / Sugar



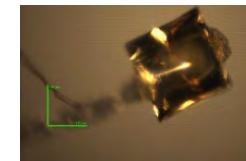
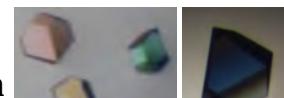
Protein X-ray crystallography



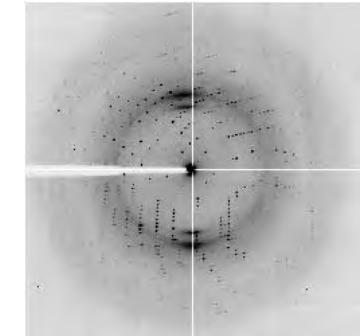
Expression
Purification



Crystallization



Freezing
& X-ray
diffraction



Phasing

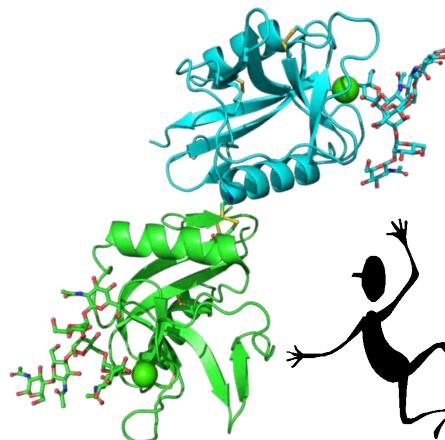


➤ Advantages

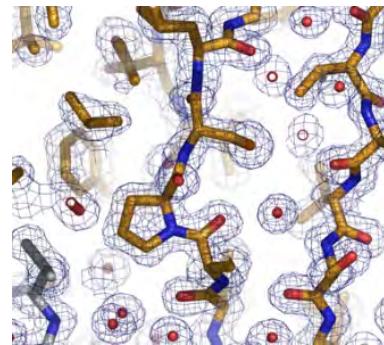
- Can go to atomic resolution
- Atomic details obtained

➤ Disadvantages

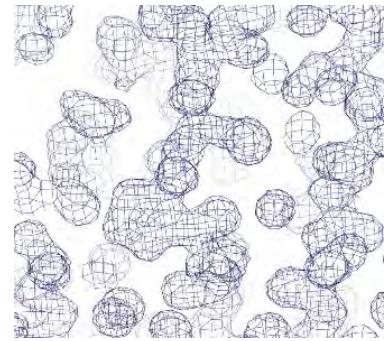
- Molecules in solid-state environment
- Require crystals
- Requires order to diffract



Refinement



Modelling

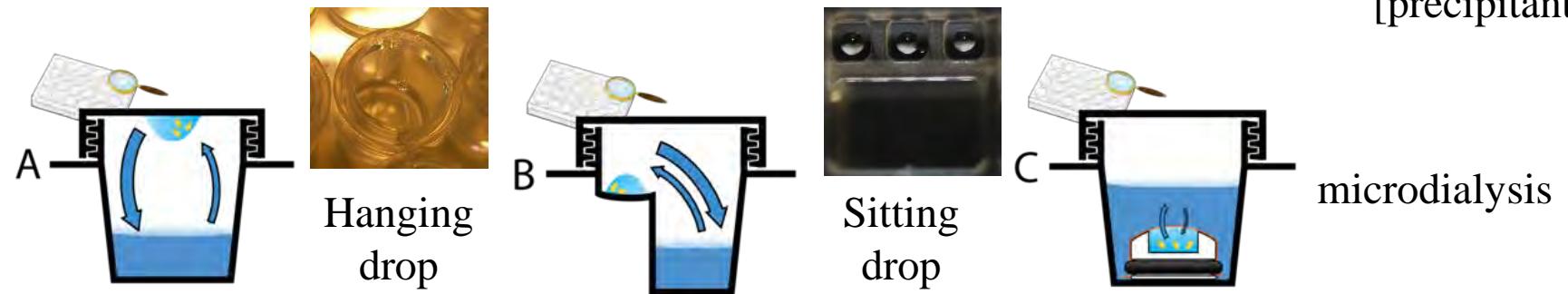




Protein crystallogenesis



- Need pure and monodisperse protein
- Empirical
 - pH, buffer, temperature, [salt], [precipitant] dependant
- Precipitants
 - High salt concentration
 - Alcohols or volatile compounds
 - Organic polymers
- Manual or robotized
- Vapor diffusion mostly used



How to obtain crystals of protein complexes with ligand

Cocrystallization



- Put the ligand in reservoir
- Preincubate with the protein
 - Limit protein dilution: >1/10
 - Set duration and temperature
 - Easily reproducible & require little ligand
- Can have crystal and spacegroup when change ligand: LecA



| Gal α 1-2Gal β 2WYF | Gal α 1-3Gal β 1- 4Glc / 2VXJ | Gal α 1-6Glc / 4AL9 | DEG144 / 4A6S | Bivalent / 4CPB | Cathecol 6YO3 | Cathecol 6YOH | Tolcapone |
|---------------------------------------|---|---|---|--|--|--|---|
| 20% PEG6K 1 M LiCl 0.1 M NaAc 4 | 10% PEG5Kmme 25 mM KSCN 0.1 M NaAc 4.6 | 20% PEG2Kmme 0.2 M KBr 0.1 M NaAc 4.6 | 0.8 M Li ₂ SO ₄ 0.1 M NaAc 4.5 | 10 % PEG8K 10% PEG1K 0.2 M MgCl ₂ 0.1 M Tris 8.5 | 20% PEG6K 1 M LiCl 0.1 M NaAc 4.5 1% DMSO | 20% PEG6K 1 M LiCl 0.1 M NaAc 4.5 1% DMSO | 24% PEG2Kmme 0.1M KSCN 0.1M NaAc 4.5 |
| 2.4 Å, P2 ₁ | 1.9 Å, P1 79.2 86.5 119.1 93.9 98.2 90.1 | 1.75 Å, P1 50.1 58.1 75.9 101.1 92.9 101 | 2.15 Å, P3 ₂ 1 | 2.15 Å, P2 ₁ 2 ₁ 2 ₁ | 1.84 Å, I2 | 1.84 Å, P2 ₁ 22 ₁ | 1.3 Å, C222 ₁ |
| | | | | | | | |

Soaking



➤ Dry soaking

- Let 0.1-1 μ L ligand solution to dry before making drop

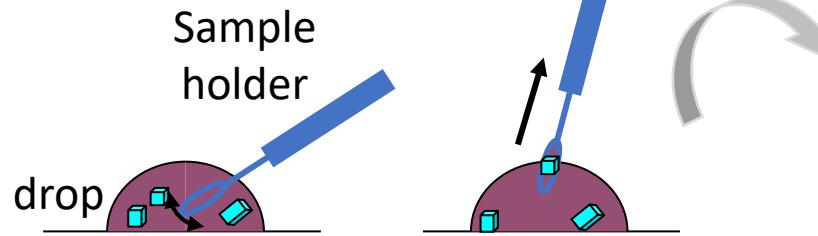
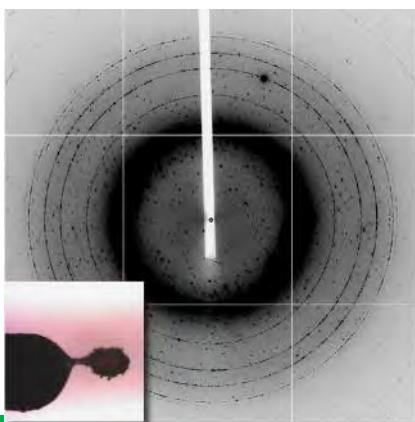
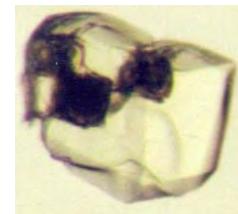
➤ Add the ligand to the reservoir solution

- Add concentrated solution or a bit of powder
- Transfer the crystal in solution with ligand
 - Try different concentrations and soaking time

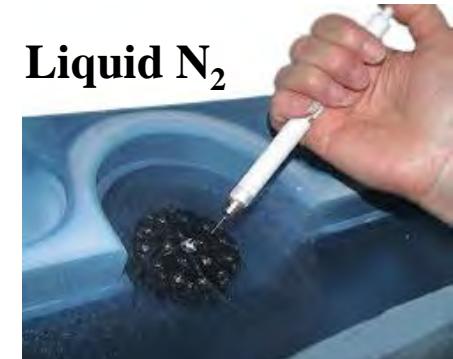


➤ Soaking whilst freezing

- Freezing limits radiation damages
- Add ligand to the cryoprotectant solution
 - Less chance of replacement by glycerol, ethylene glycol and MPD



Liquid N₂





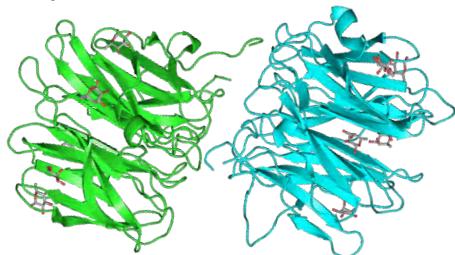
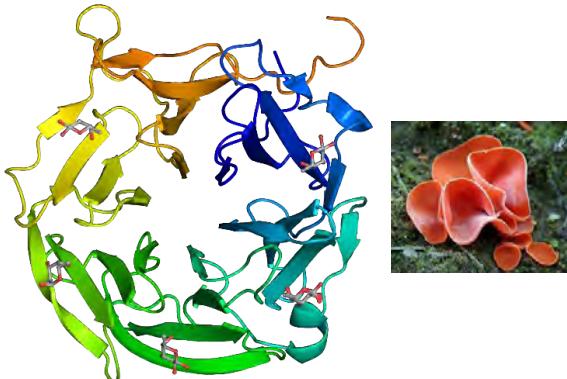
Phasing: molecular replacement



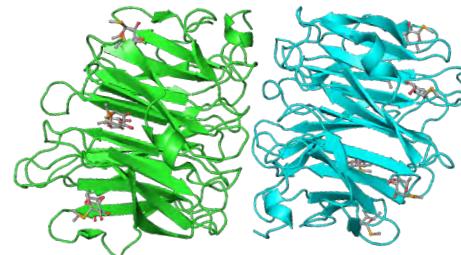
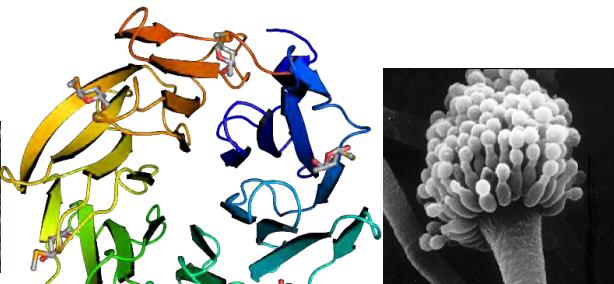
➤ Have a good model from homologous protein

- Sequence identity > 25%
- Can use alphafold if in Uniprot or fold known
- Good conservation secondary structure: difficult for β -sheets

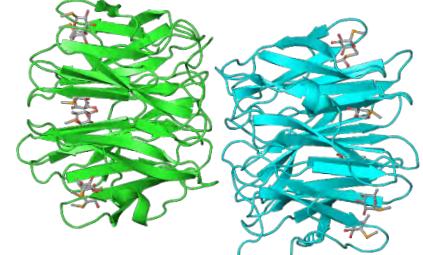
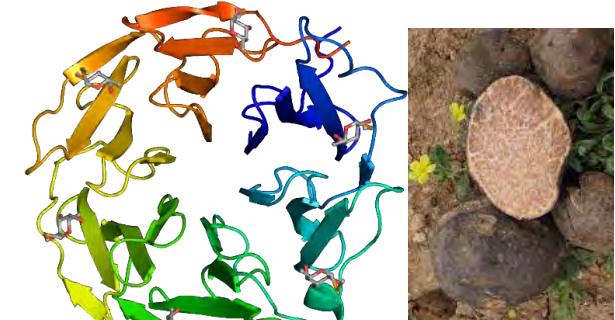
AAL/1OFZ/Hg
Aleuria auratia



FleA/4AGI/SFU (35%)
Aspergillus fumigatus



TrfbL1/SFU (32/43%)
Terfezia boudieri





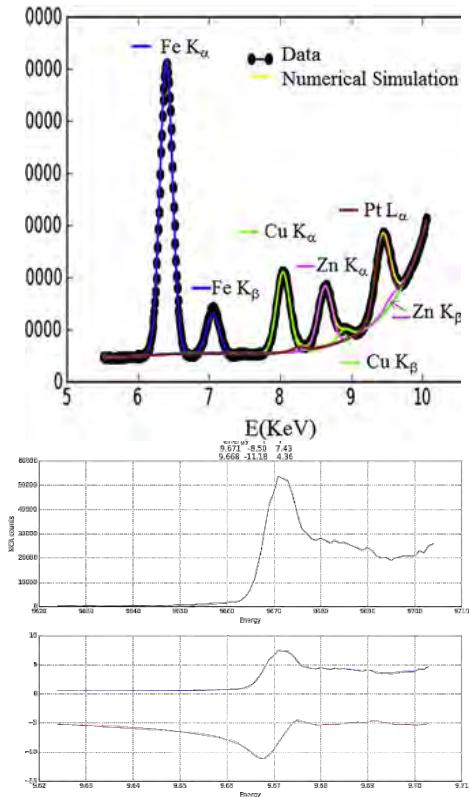
Phasing: isomorphous or anomalous



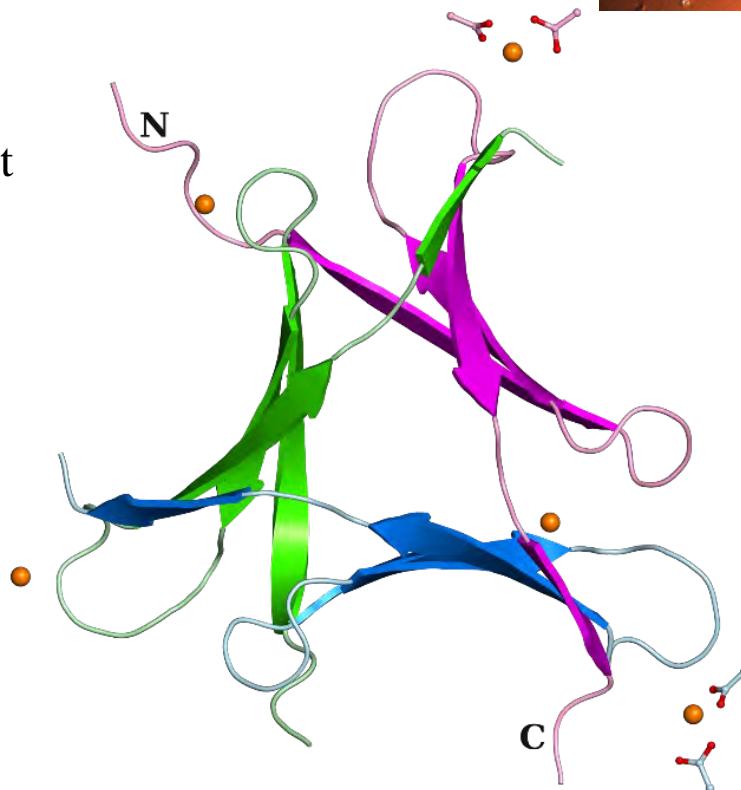
➤ Look in your crystallization conditions

- PhoSL from *Pholiota squarrosa*

- 300 mM Zinc acetate, 0.1 M Imidazole-HCl pH 6-7



X-ray fluorescent
XRF scan

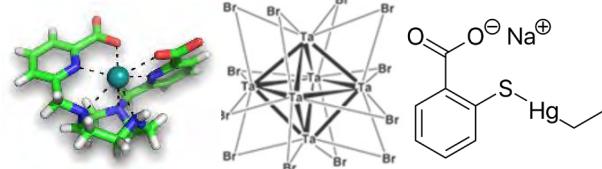




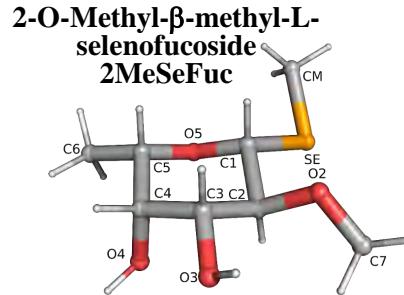
Phasing: isomorphous or anomalous-2



- Modified your protein with selenomet or selenocys
- Use heavy metals, clusters of lanthanides
- Use ligand with heavy atoms (Se, Br, S, F)



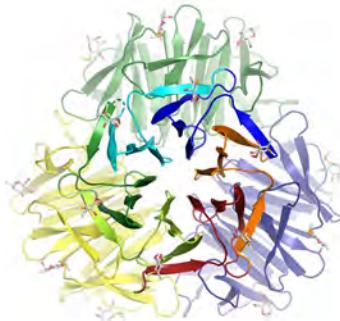
Lb-Tec2



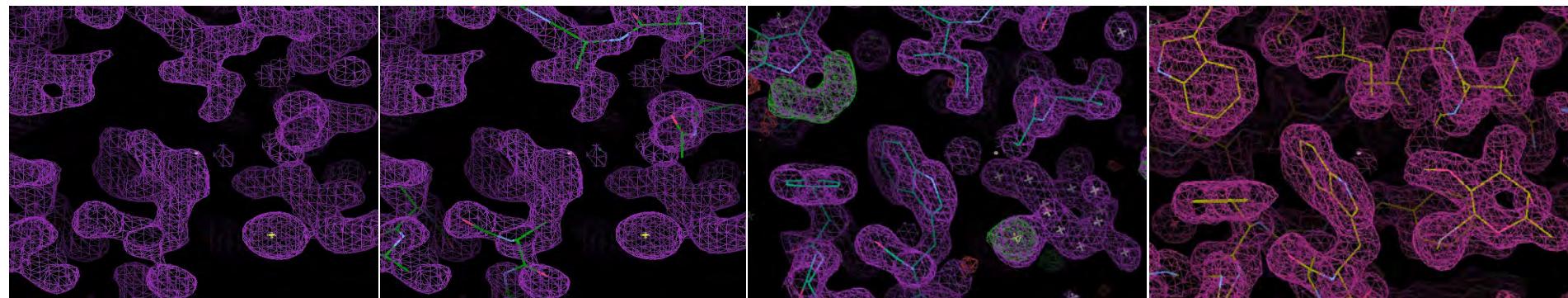
Se peaks



Alex Titz
HIPS
Saarbrücken
Germany
R Sommer



Final



Carbohydrate refinement



➤ Find proper 3-letter code

- Check ligand database <http://ligand-expo.rcsb.org/ld-search.html>

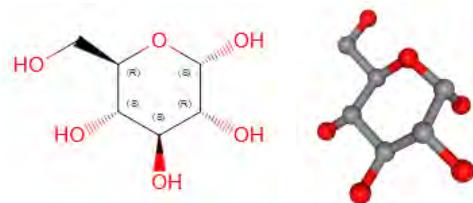


Ligand Expo Search Result Summary

Query: glucopyranose

Query type: Molecular name (exact sub-string)

Result count: 252



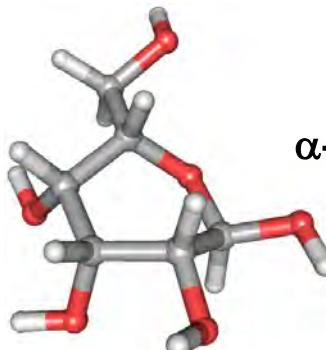
GLC

alpha-D-glucopyranose

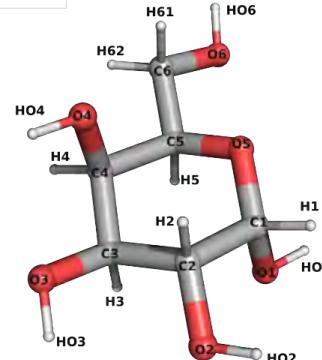
Find entries where: GLC

- is present as a standalone ligand in 301 entries
- as a non-polymer is covalently linked to polymer or other heterogen groups 61 entries
- is present in a branched oligosaccharide 1,638 entries

| Chemical Component Summary | |
|----------------------------|--|
| Name | alpha-D-glucopyranose |
| Synonyms | alpha-D-glucose; D-glucose; glucose |
| Identifiers | (2S,3R,4S,5S,6R)-6-(hydroxymethyl)oxane-2,3,4,5-tetrol |
| Formula | C ₆ H ₁₂ O ₆ |
| Molecular Weight | 180.16 |
| Type | D-SACCHARIDE, ALPHA LINKING |
| Isomeric SMILES | C([C@H]1[C@H](C([C@H]([C@H]([C@H](O1)O)O)O)O)O) |
| InChI | InChI=1S/C6H12O6/c7-1-2-3(8)4(9)5(10)6(11)12-2/h2-11,1H2H2,3-,4+,5-,6-/m1/s1 |
| InChIKey | WOZGKKKJUFFOK-DVKNGEFBSA-N |
| Chemical Details | |
| Formal Charge | 0 |
| Atom Count | 24 |
| Chiral Atom Count | 5 |
| Bond Count | 24 |
| Aromatic Bond Count | 0 |



GLA
α-D-galactose



- 1 code per anomer
 - α-D-Glucose: GLC
 - β-D-Glucose: BGC
 - β-L-Glucose: Z8T

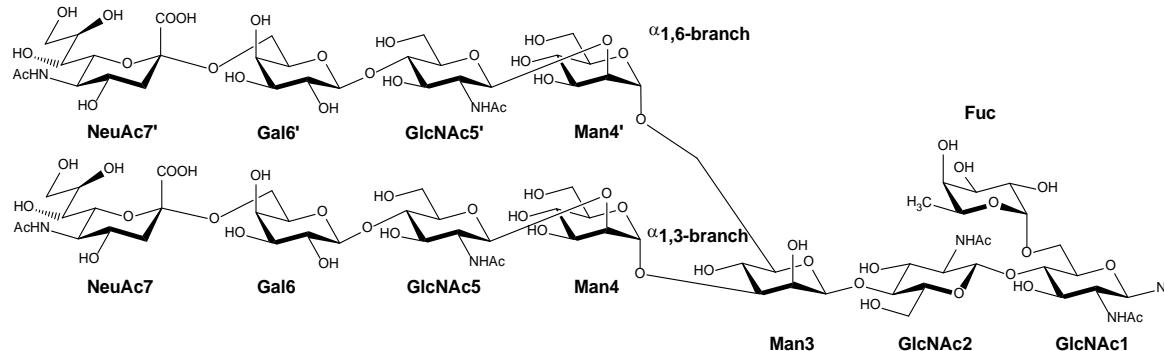
- Check library
 - No distortion



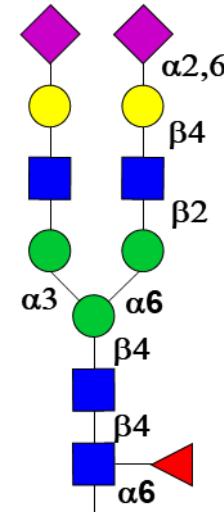
Oligosaccharide refinement



- Define correct linkage description
- Programs do not know how to deal with L-sugars



6FX3

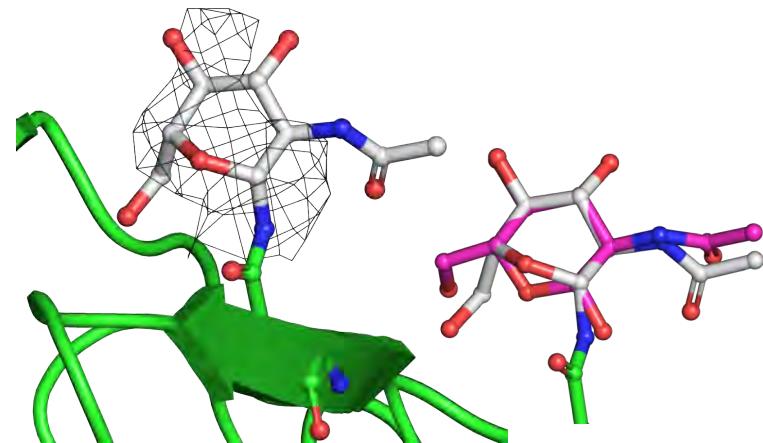
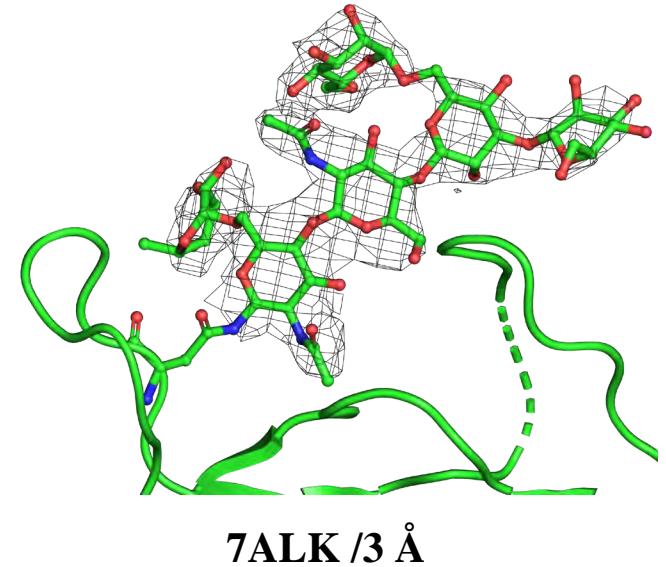
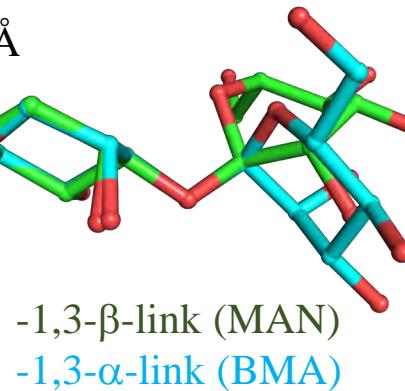
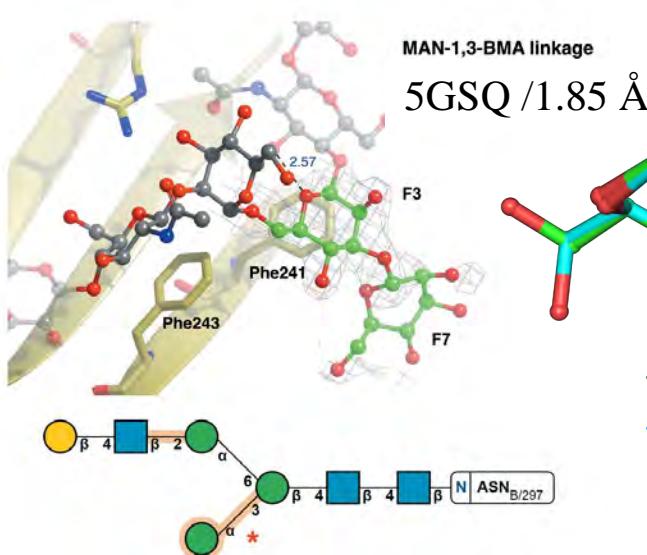


| | | | | | | | | | |
|-------|----|-----|---|-----|----|-----|---|-----|----------|
| LINKR | 06 | C4W | A | 102 | C1 | FUC | A | 101 | BETA1-6 |
| LINKR | 04 | C4W | A | 102 | C1 | NAG | A | 103 | BETA1-4 |
| LINKR | 04 | NAG | A | 103 | C1 | BMA | A | 104 | BETA1-4 |
| LINKR | 06 | BMA | A | 104 | C1 | MAN | A | 105 | ALPHA1-6 |
| LINKR | 02 | MAN | A | 105 | C1 | NAG | A | 106 | BETA1-2 |
| LINKR | 04 | NAG | A | 106 | C1 | GAL | A | 107 | BETA1-4 |
| LINKR | C2 | SIA | A | 108 | 06 | GAL | A | 107 | SIA-GAL |
| LINKR | 03 | BMA | A | 104 | C1 | MAN | A | 109 | ALPHA1-3 |
| LINKR | 02 | MAN | A | 109 | C1 | NAG | A | 110 | BETA1-2 |

Carbohydrate validation



- Do not overfit at low resolution
- Check nomenclature, ring conformation, density fit
 - Privateer
 - PDB-REDO



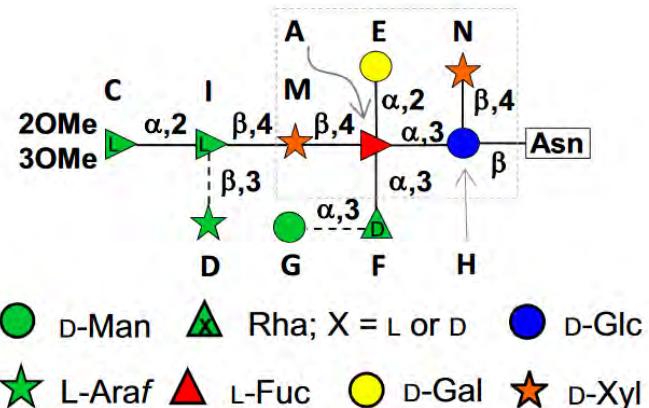


Tricky for non structural glycobiologists

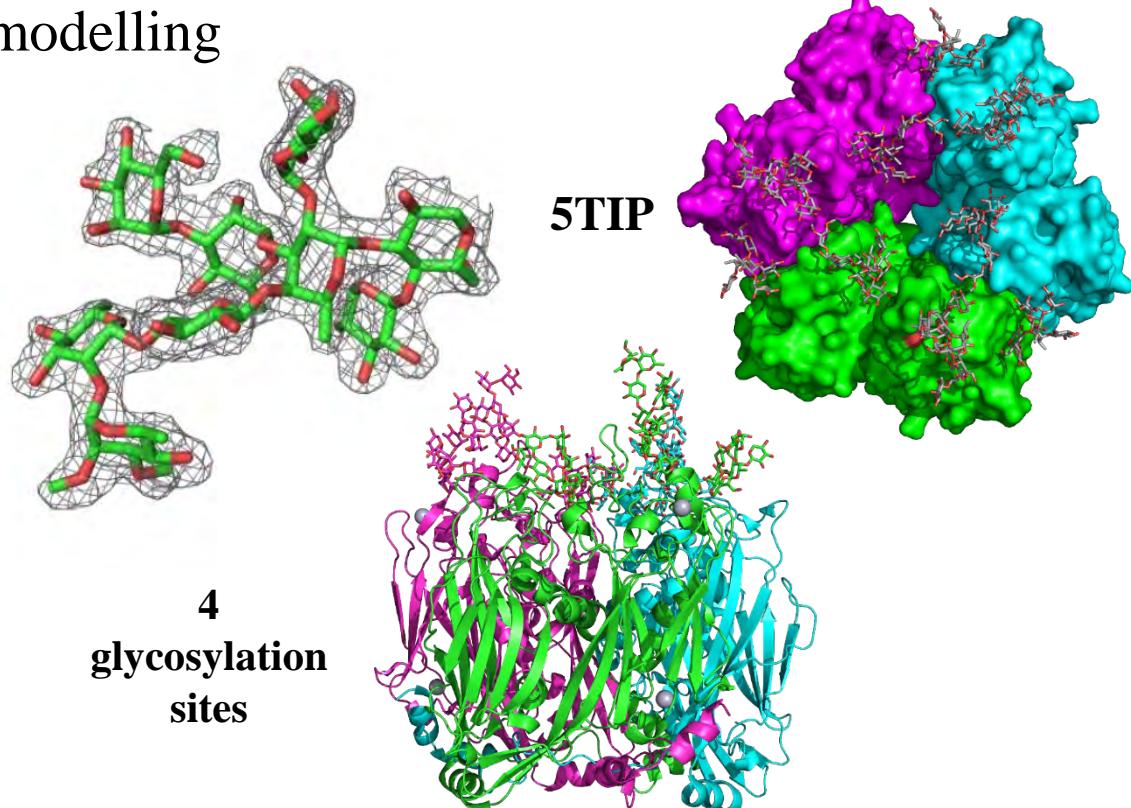


► Major capsid protein (Vp54) of chlorovirus PBCV-1

- X-ray 2002: try to fit classical N-glycans
- Sugar NMR 2013: highly complex N-glycosylation
- X-ray revised in 2017 + modelling



H = BGC **A** = FUC
N/M = XYP **E** = GLA
F = XXR **G** = MAN
I = RM4 **C** = 7CV



4
glycosylation
sites

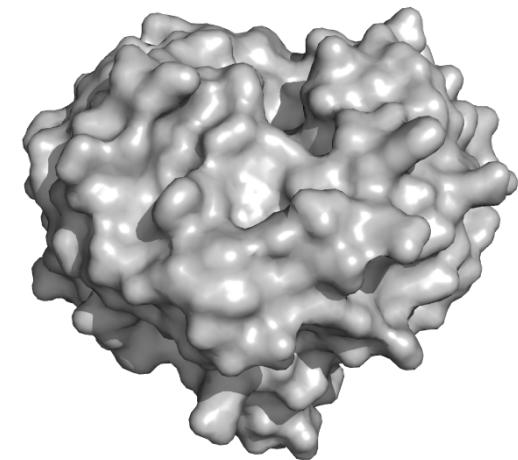
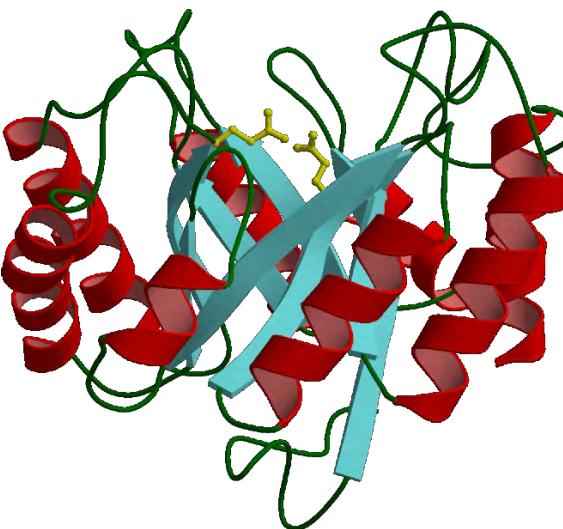
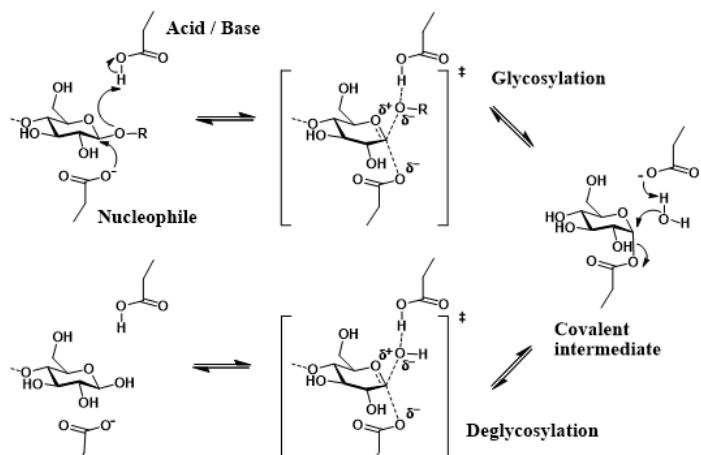
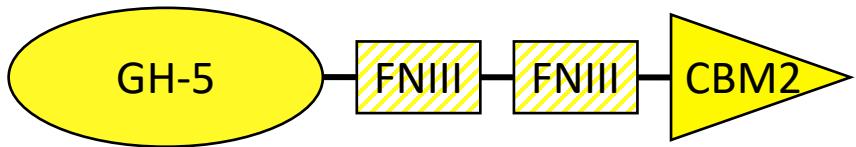


Insights in the retaining mechanism of GHs



➤ Cellulase Cel5A from *Bacillus agaradhaerens*

- Modular endoglucanase
- Active pH range 5.0-13.0
- Glu139 (acid/base) and Glu228 (nuc)

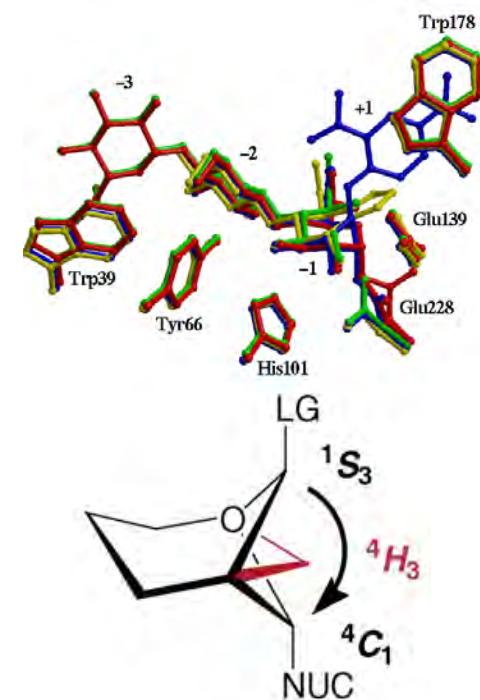
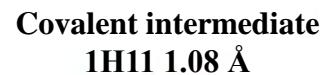
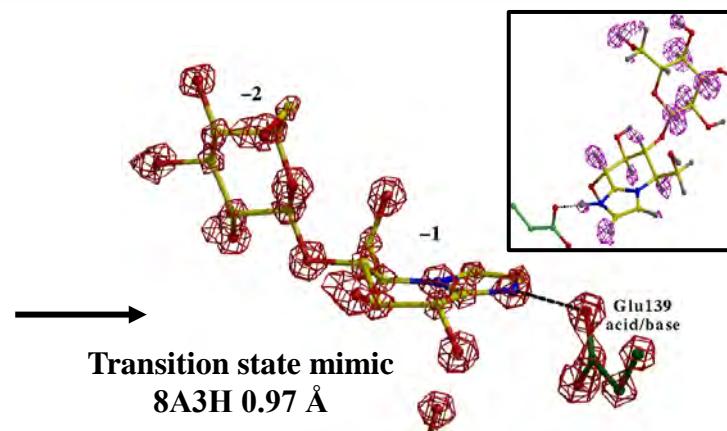
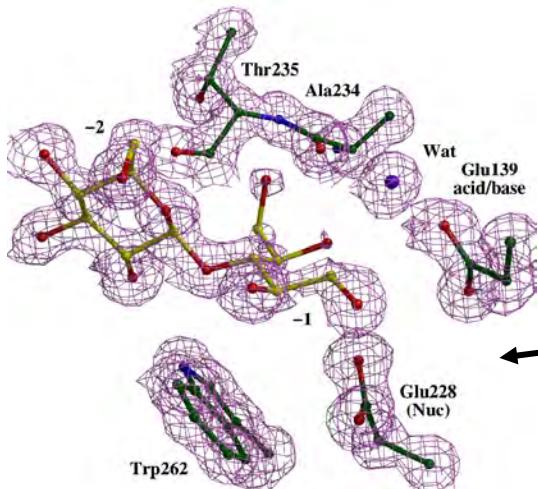
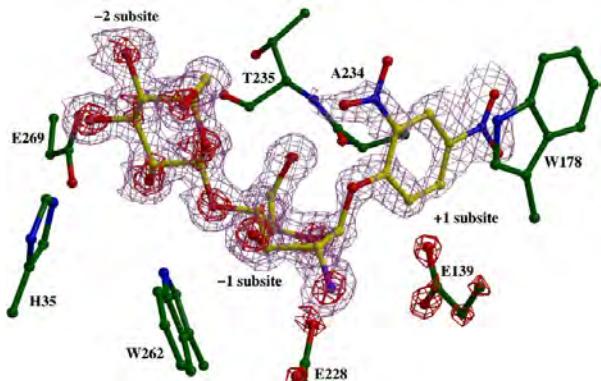


➤ Trapping of each step by X-ray crystallography

- Use specific ligand, mutated protein, inactive pH



Snapshots at atomic resolution





GHs inhibitors design

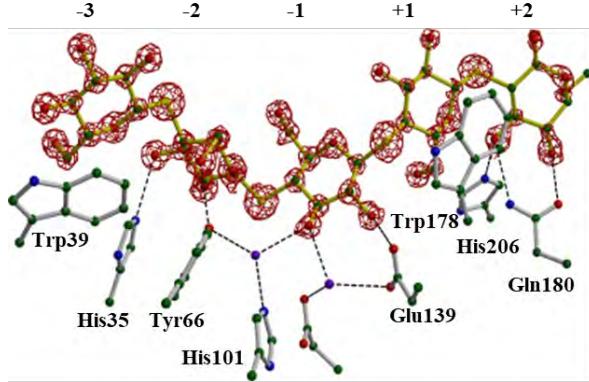


➤ Interconversion of sugar ring conformations

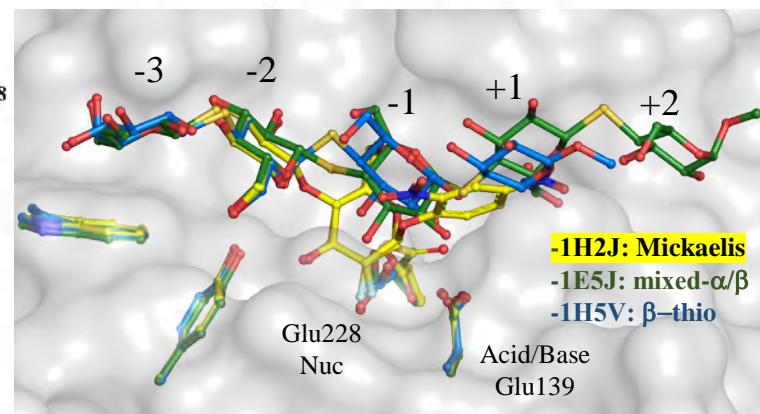
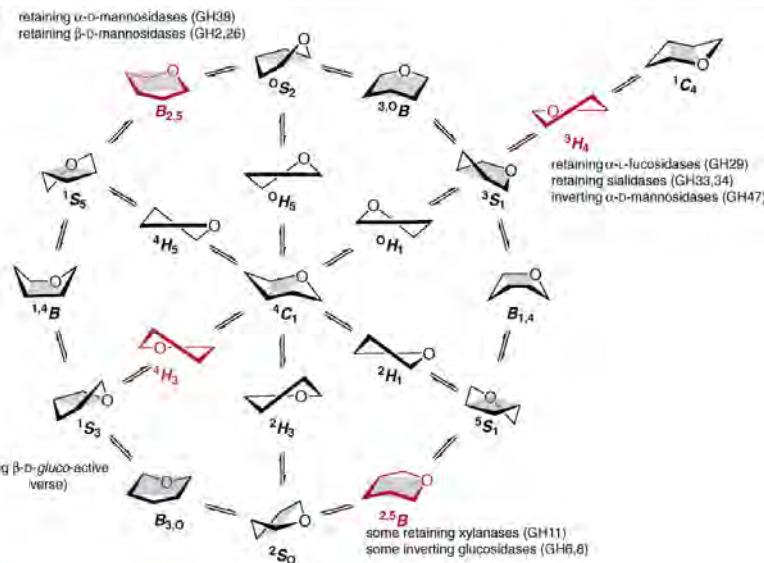
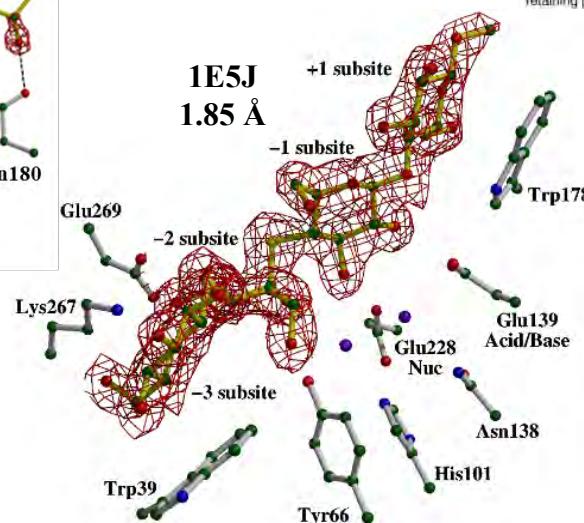
- Highlights transition state
- Essential for inhibitor design

➤ Serendipity

- New inhibitors class for glucosidases



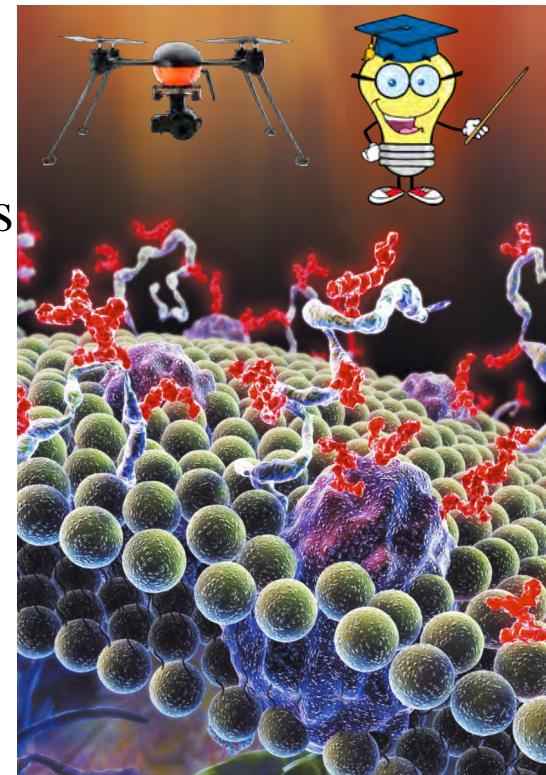
1H5V
1.1 Å



Lectins



- Ubiquitous carbohydrate binding proteins
- Specific and reversible sugar binding without modification
- Decipher the glycocode
- Implicated in many cellular processes
 - From warning their kin to poisoning their enemies
- Multivalency compensates for low affinity
 - Give ability to agglutinate cells
- New database: <https://unilectin.unige.ch/>

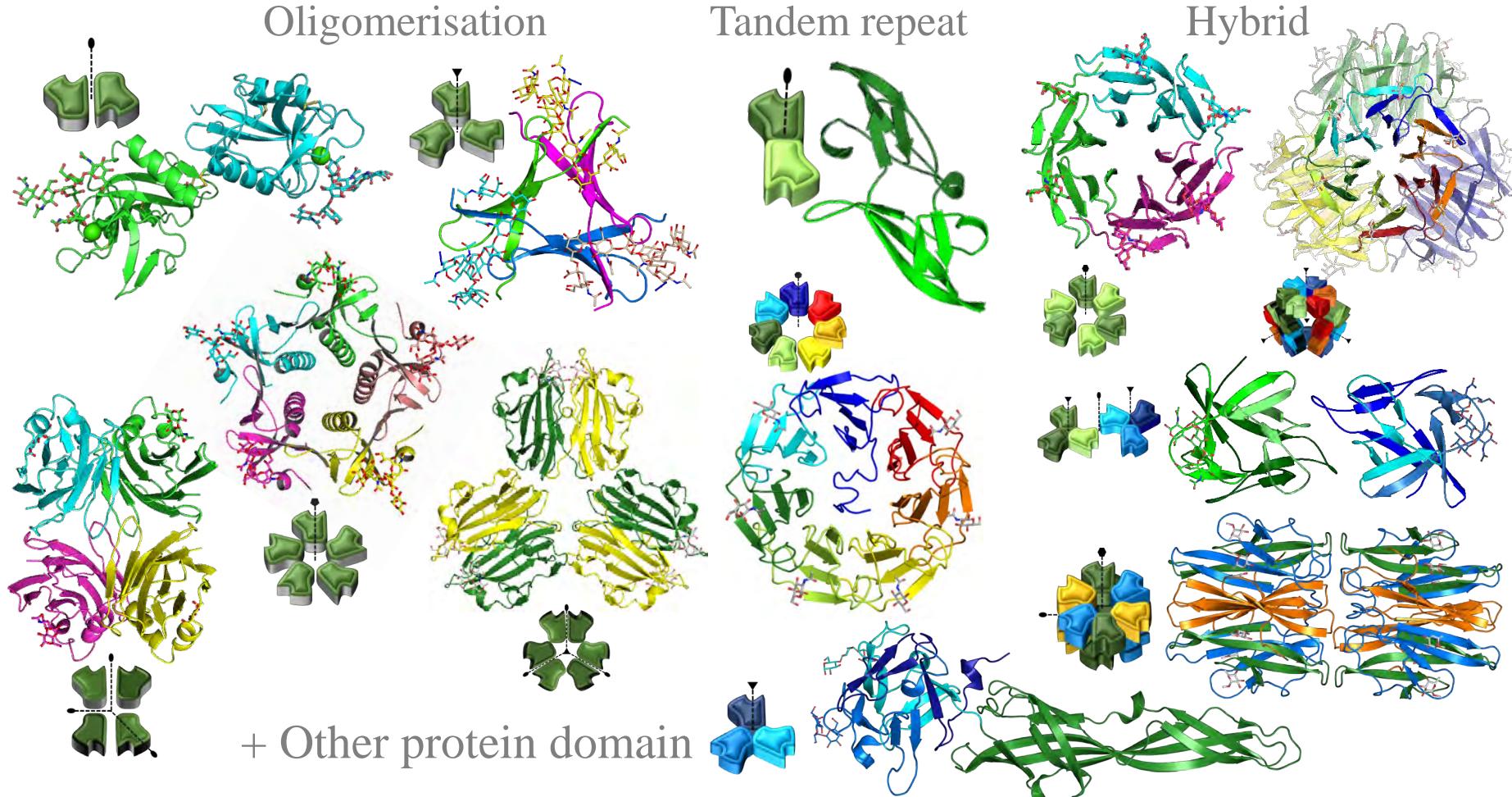




Lectin structure



➤ High diversity of fold and quaternary structure

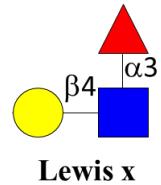
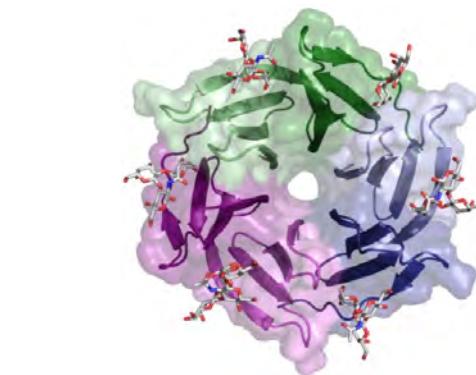




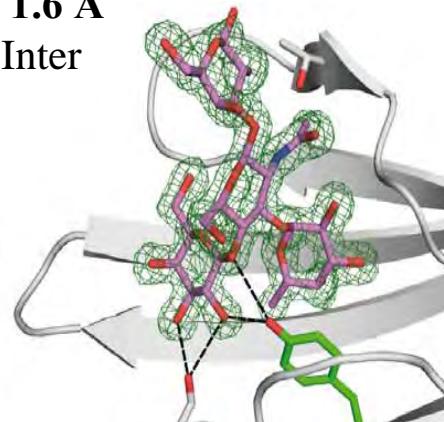
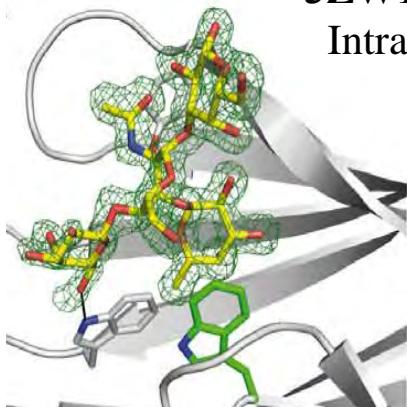
Rare conformation



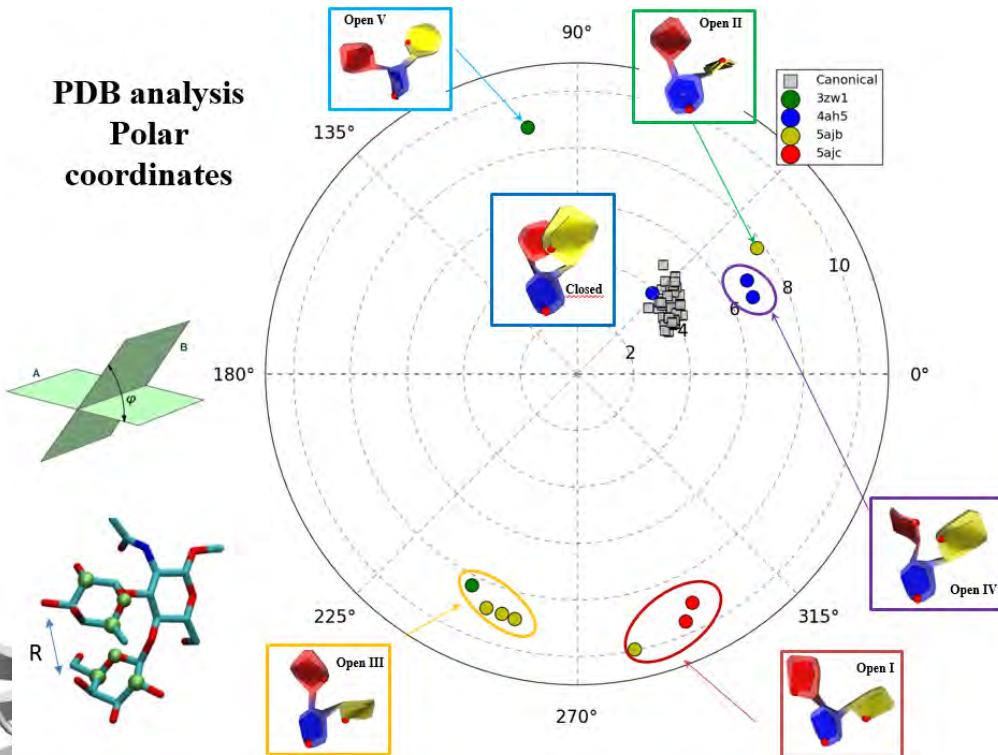
- 1 structure of lectin complex with ring distortion
- BambL/RSL: Hidden conformation of Lewis X



3ZW1, 1.6 Å
Intra/ Inter



PDB analysis
Polar
coordinates



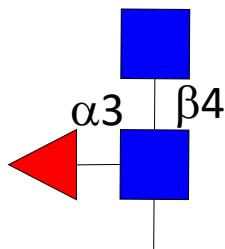


Multivalency and function

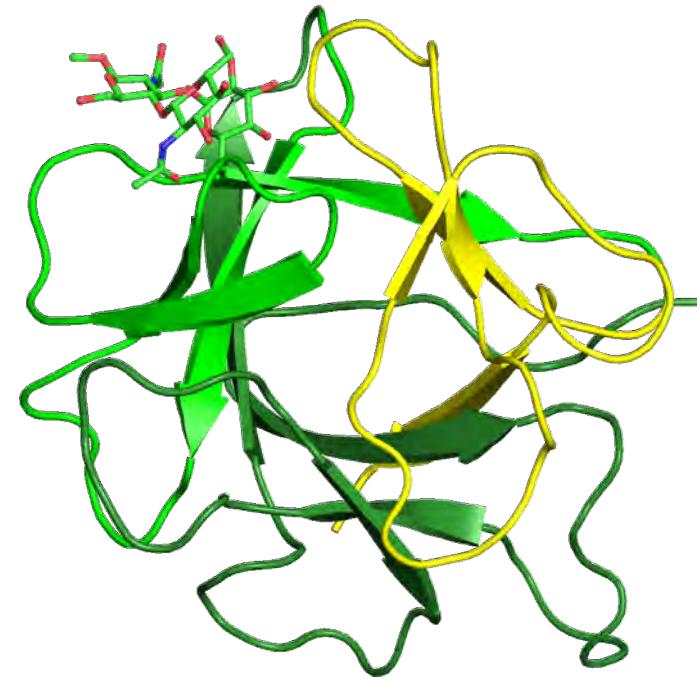


➤ CCL2 from *Coprinopsis cinerea*

- Involved in innate immunity and defense
- Toxic for nematodes and flies
 - Recognised 3-core fucose of midgut N-glycans



- Monomer on Superdex75
- RMN structure: β -trefoil fold
 - Only subsite β functional
- Could not explain toxicity
 - Monomer and monovalent



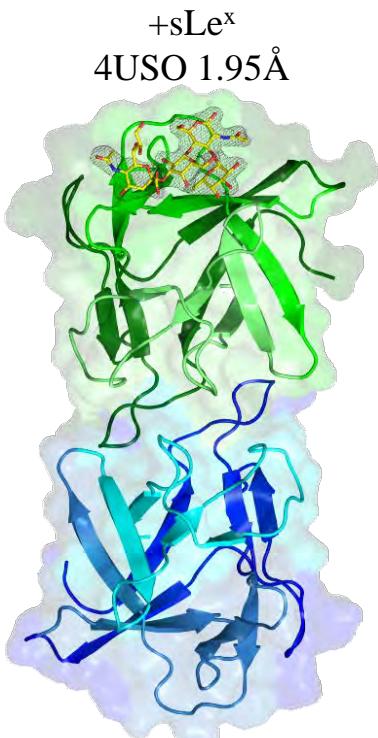
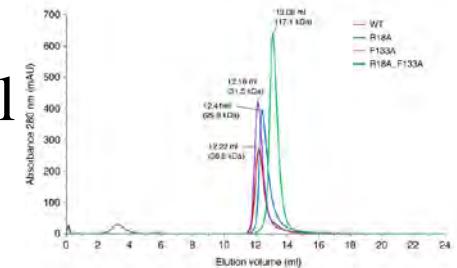
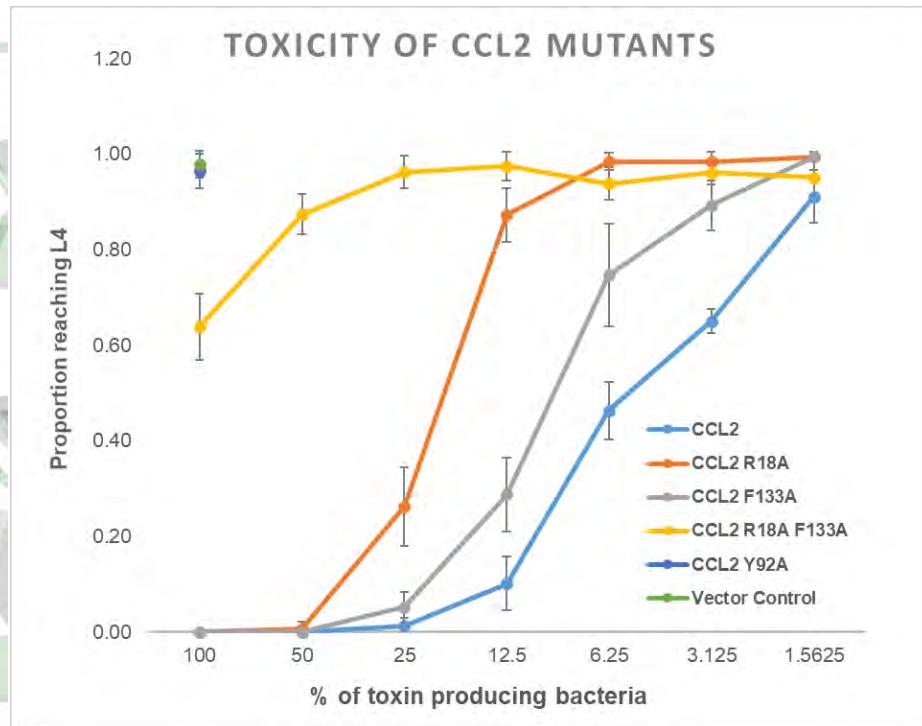
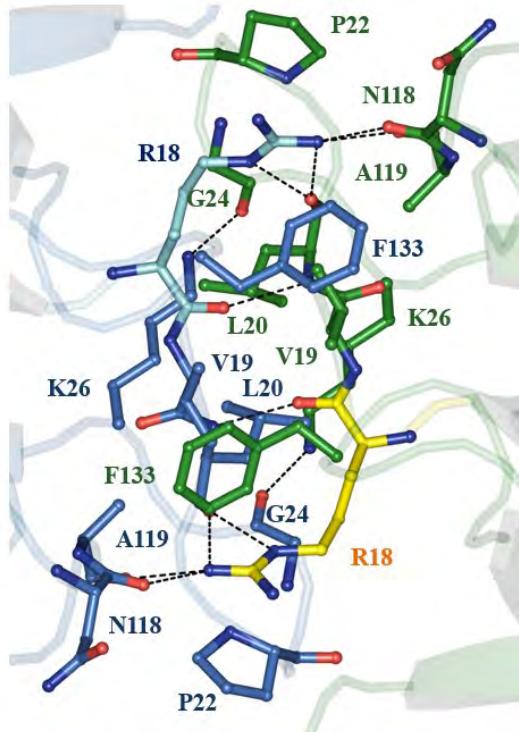


Multivalency and function-2



► CCL2 is a dimer

- Confirmed by SEC on Enrich70, DLS, Native gel
- Toxicity dependent on its dimerization
 - Disruption of dimer interface requires double mutation

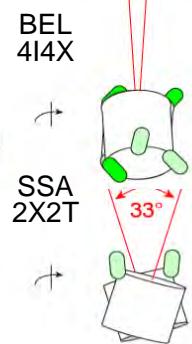
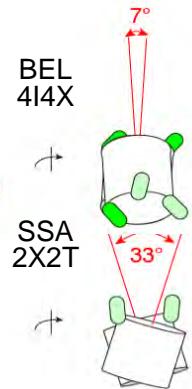
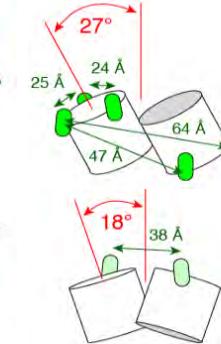
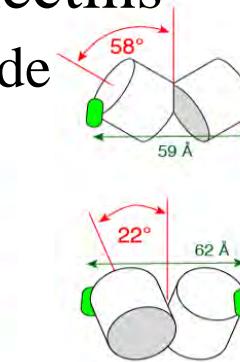
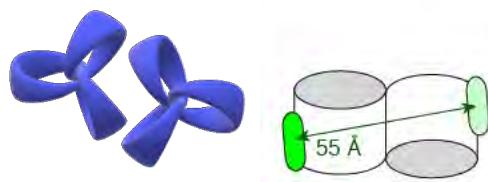




Multivalency and function-2



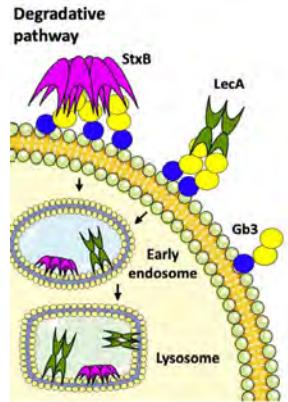
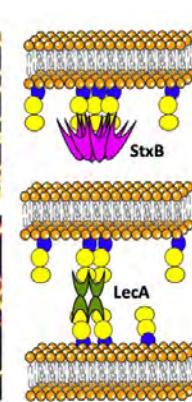
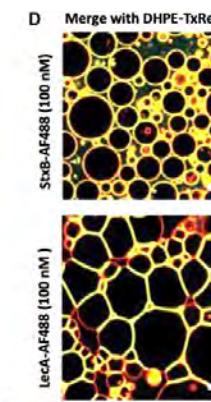
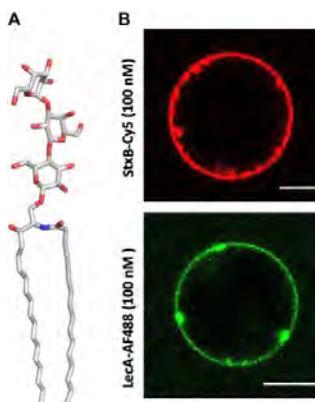
- Spatial distribution of binding sites impact function and recognition mode of β -trefoil lectins
 - CCL2: 1 binding surface on the side



- Others: 1/2 binding surfaces on the top

- Reorganisation of membrane glycoconjugates

- Clustering of glycolipids
- Crosslinking
- Change in membrane dynamics
- Endocytosis

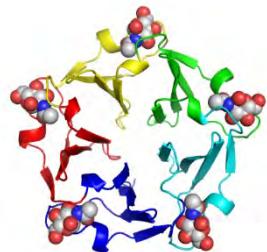




Lectins with β -propeller fold



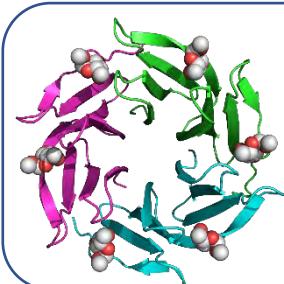
Tachylectin 2
GlcNAc



*Tachipileus
tridentatus*

Beisel *et al.*
EMBO J. 1999

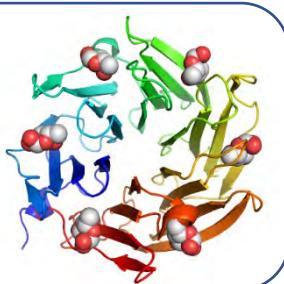
RSL
fucose



*Ralstonia
solanacearum*

Kostlanova *et al.*
J. Biol. Chem. 2005

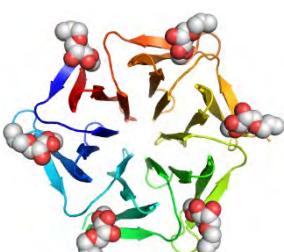
FleA
fucose



*Aspergillus
fumigatus*

Houser J, *Plos One*,
2013, 8:e83077

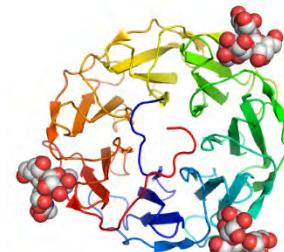
Tectonin/Lb-Tec2
Me-sugar



*Laccaria
bicolor*

Sommer *et al.*
Structure 2018

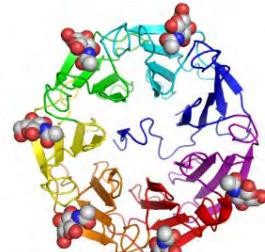
PLL
fucose



*Photorhabdus
luminescens*

Kumar *et al.*
J. Biol. Chem. 2016

PVL
GlcNAc



*Pstatyrella
velutina*

Cioci *et al.*
J. Mol. Biol. 2006

➤ Define blade signature for prediction: PropLec in Unilectin

PropLec5A



PropLec6A



PropLec6B



PropLec7A



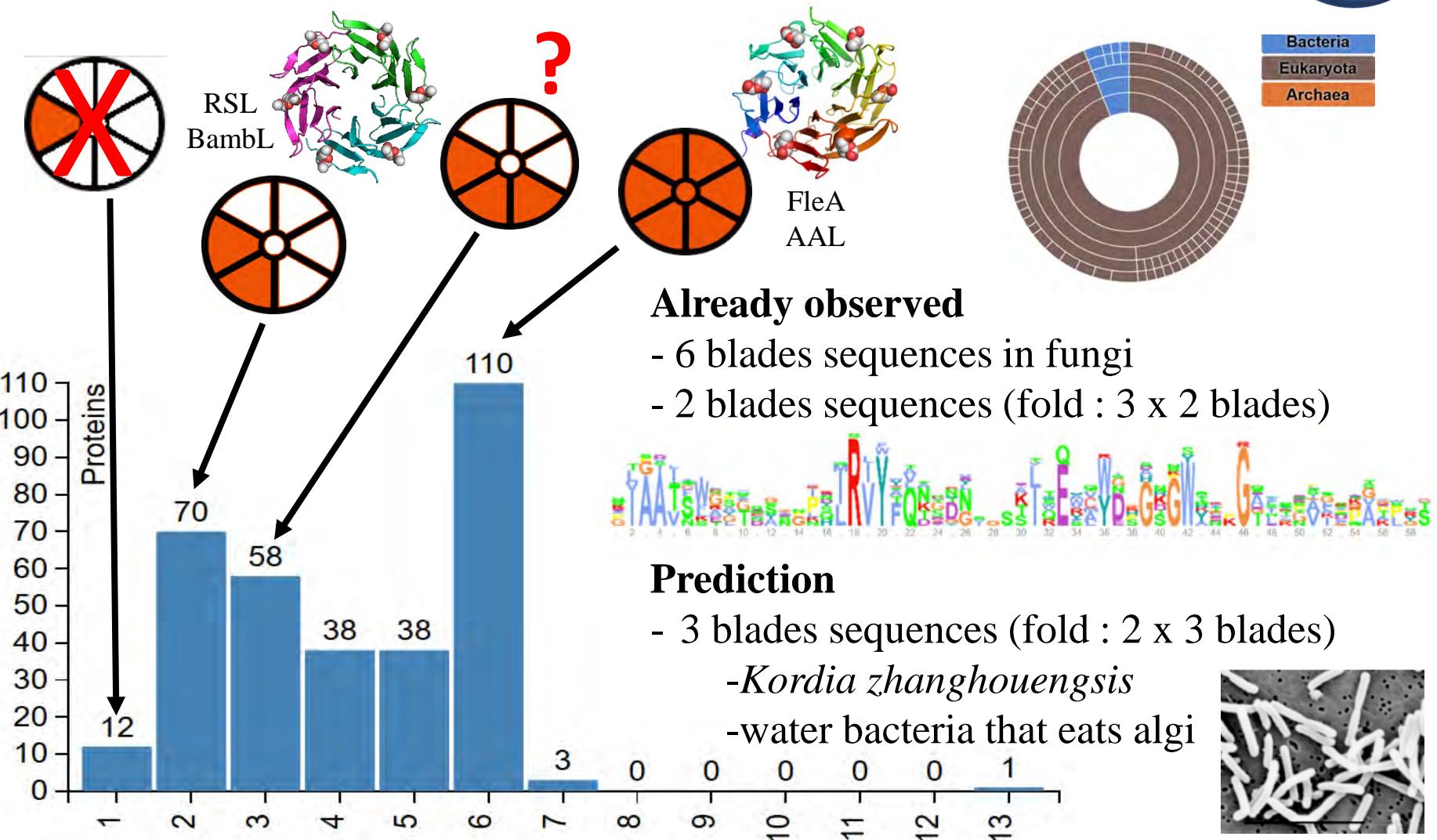
PropLec7B



PropLec7C



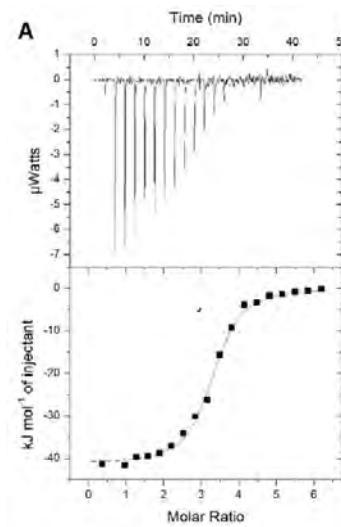
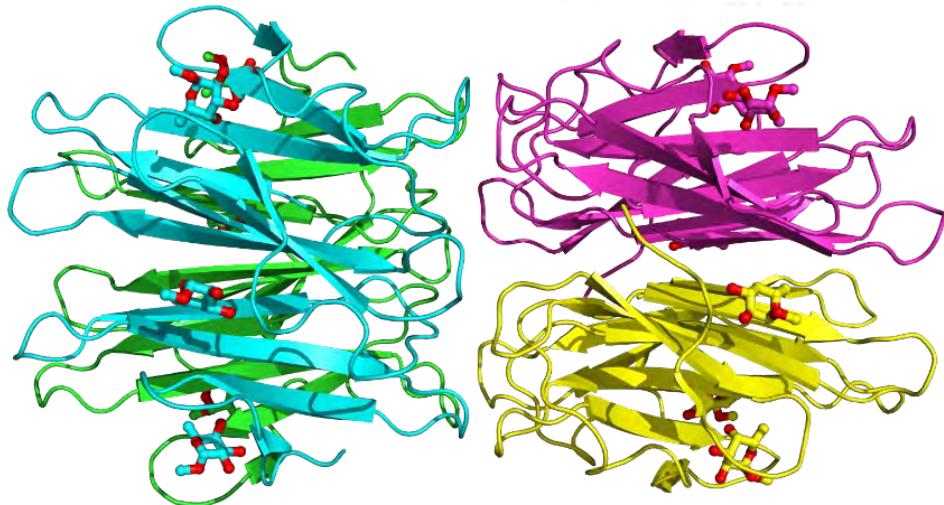
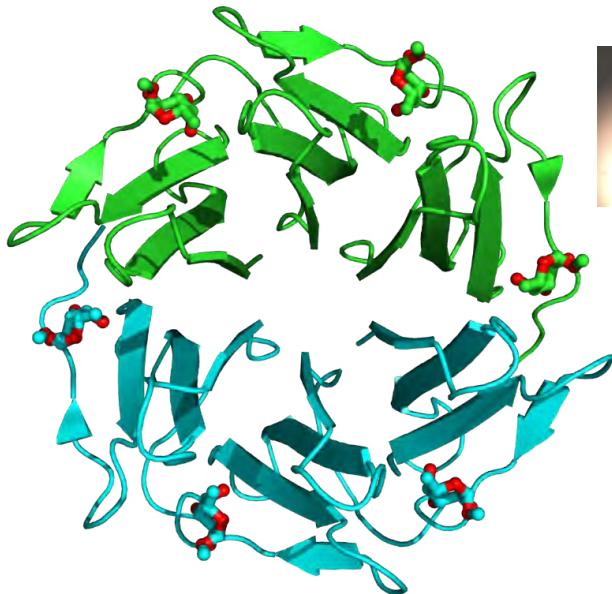
Distribution of predicted blade in PropLec6A



Recombinant KozL



- Overexpressed in *Escherichia coli*
 - Purified on mannose agarose: 35 mg/L
 - ITC: 3 fucose binding sites
 - Tetramer by AUC
- Structure solved by SAD using α SeMeFuc
 - **Prediction corroborated:** 2 x 3 blades

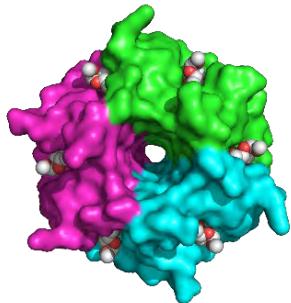




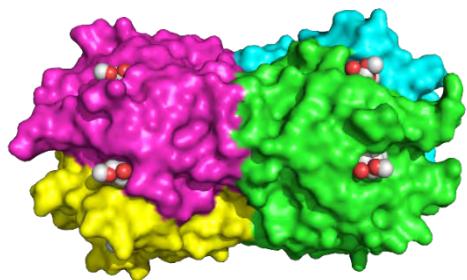
PropLec6: many arrangements



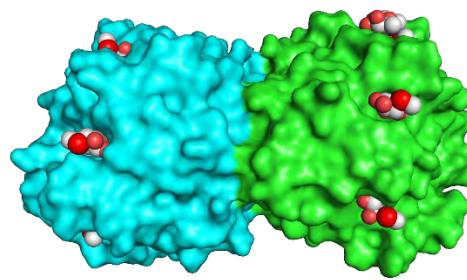
BambL/RSL
6 Fuc



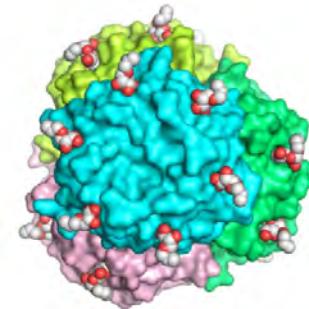
KhozL
12 Fuc



FleA/ SapL1
12 Fuc

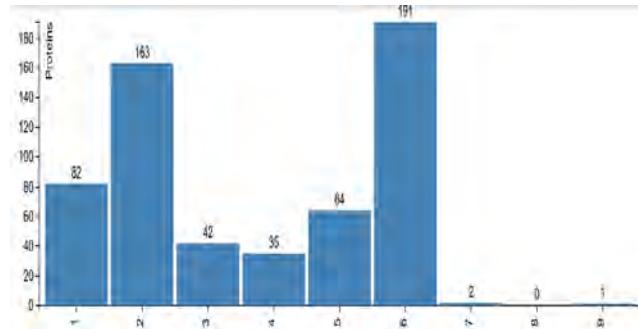


Lb-Tect2
24 Me-sug



➤ Is 1 blade occurring?

- Fold: 6x1 blade

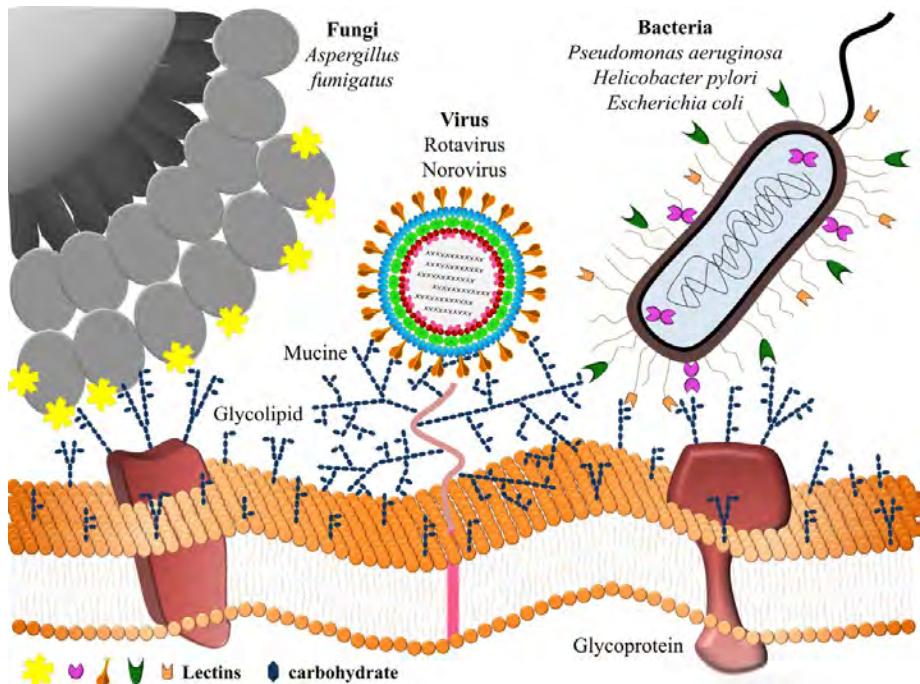




Lectins as therapeutic targets



- Lectins used host glycoconjugates as entry points
 - Mediate host recognition and adhesion
 - Blocking lectins → prevent infections



Classic:
Destroy pathogens
→ selective pressure
driving **resistance**

vs
Alternative:
Disable pathogens
→ **Resistance not required for survival**
Antiadhesion therapy

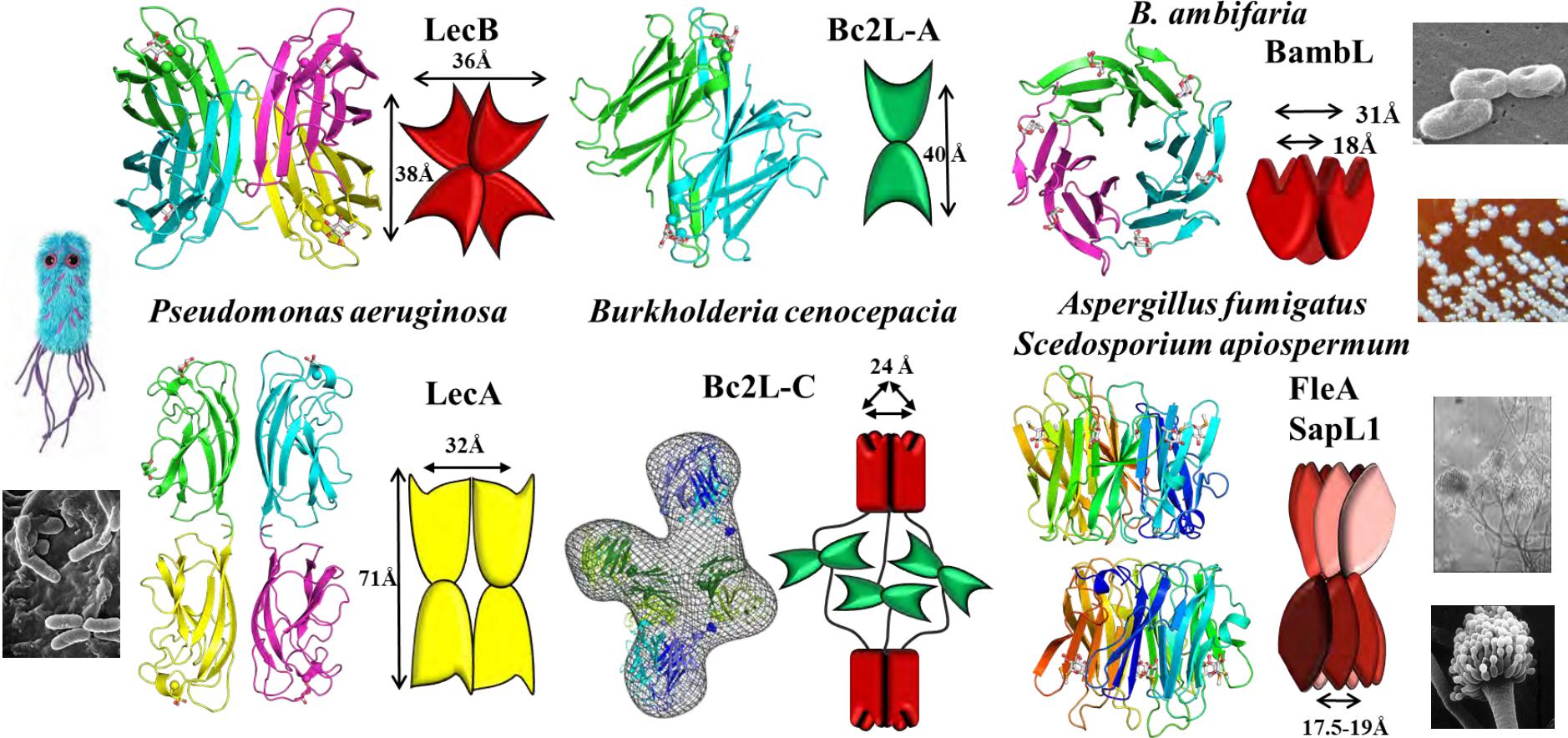
→ Development of glycocompounds as antimicrobials



Lectin targets



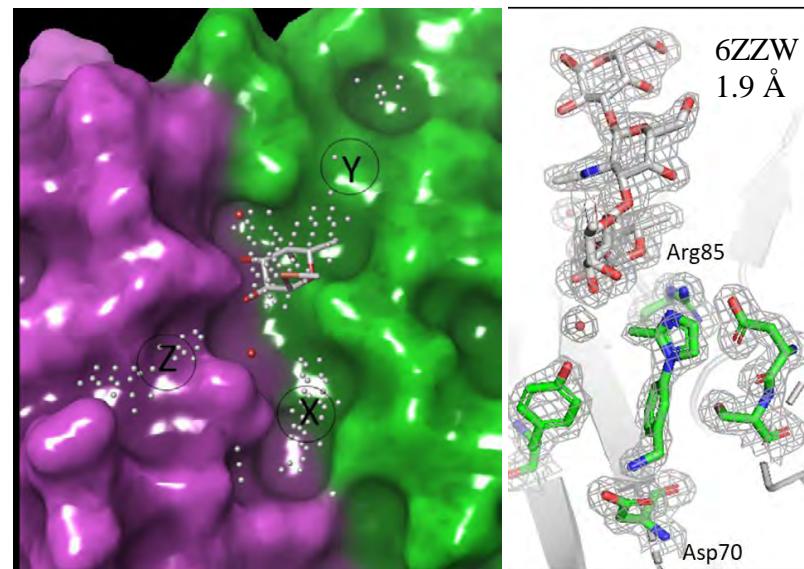
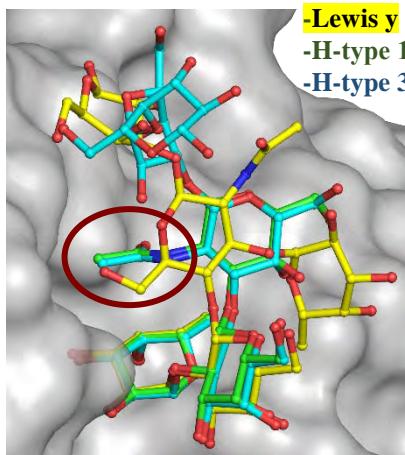
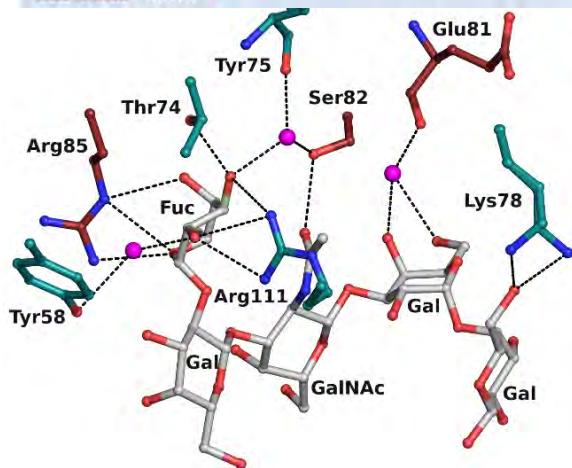
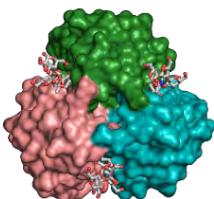
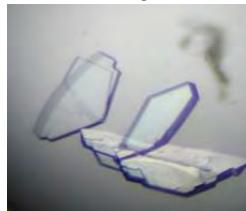
➤ Opportunistic pathogens → bronchopulmonary infections





- New construct 1-132 in pCold-TEV: good yield & stable
- Complex with oligos by cocrysts and with inhibitors by soaking

| Ligand | k_D (μM) | PDB | Res (\AA) |
|--------------------------|-------------------------|-------|----------------------|
| L-Fuc | 2430 | 2WQ4 | 1.42 |
| L-Gal | 2000 | | |
| Disac | 2500 | | |
| H-type 1 tri | 25.4 | | |
| H-type 1 tetra | 56.6 | 6TID | 1.6 |
| Lewis Y penta | 53.2 | 7TOLU | 1.6 |
| H-type 3- GloboH hexa | 26.7 | 6TIG | 1.9 |



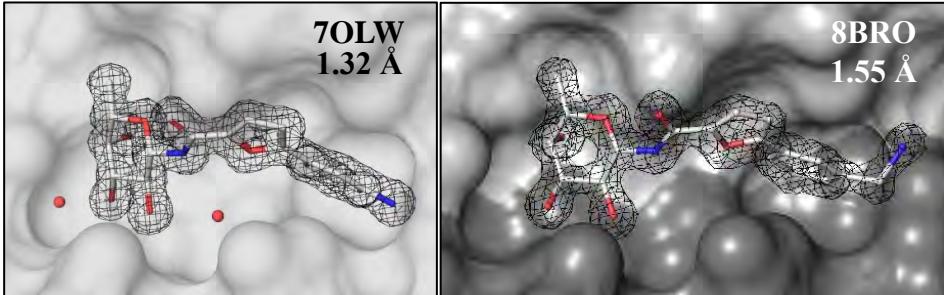
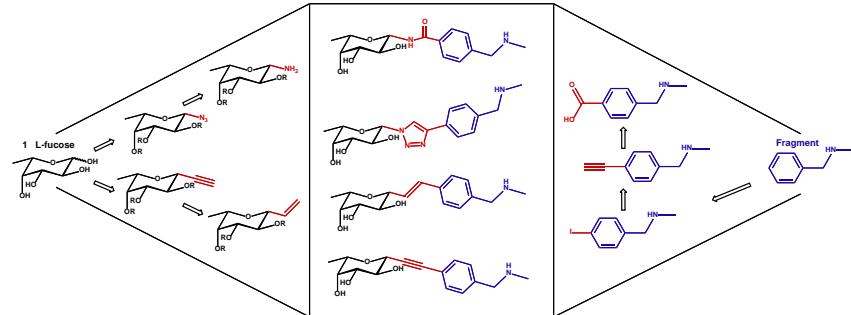
- 3 druggable sites predicted with SiteMap in Maestro



Bc2LC-nt inhibitors

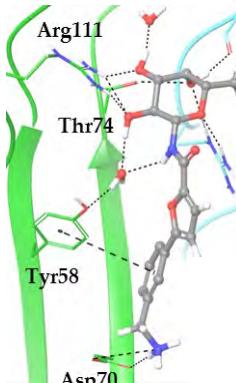


► Design, synthesis & evaluation of fucosides derivatives

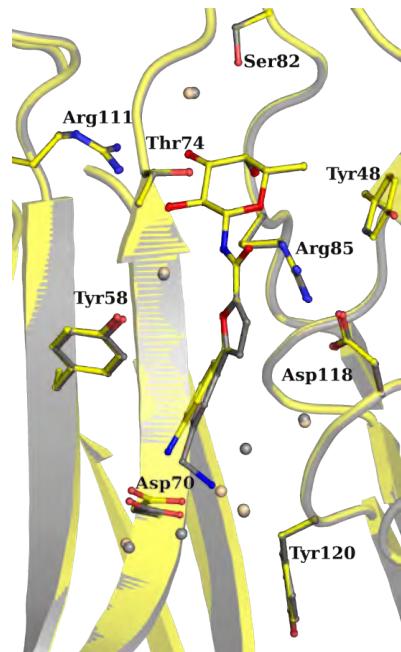
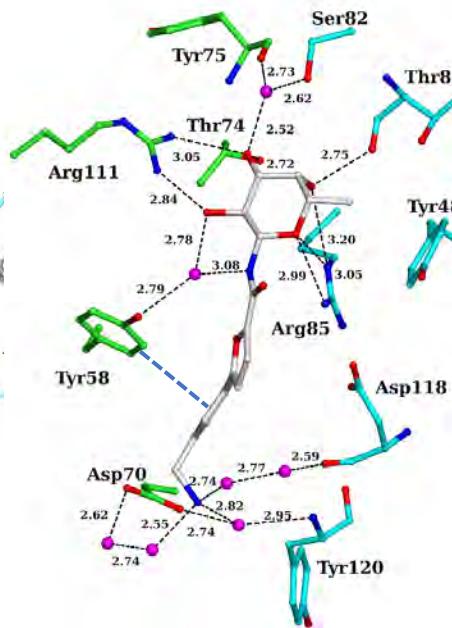


- Transform aniline in aminomethylene
 - One order of magnitude gain
 - Terminal amine does indirect Hbonds

| Ligand | k_D (μM) | PDB | Res (\AA) |
|--------|----------------------------|------|-------------------------|
| L-Fuc | 2700 | | |
| 22a | 280 | 7OLU | 1.6 |
| 8c | nd | 7OLW | 1.32 |
| 3 | 159 | 8BRO | 1.55 |
| 4 | 390 | | |



Modelling



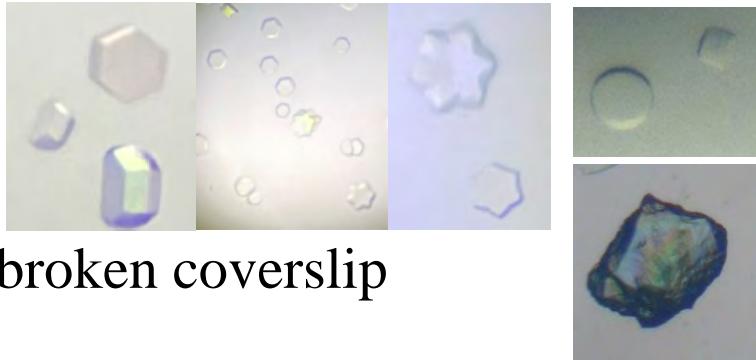


Miscellaneous



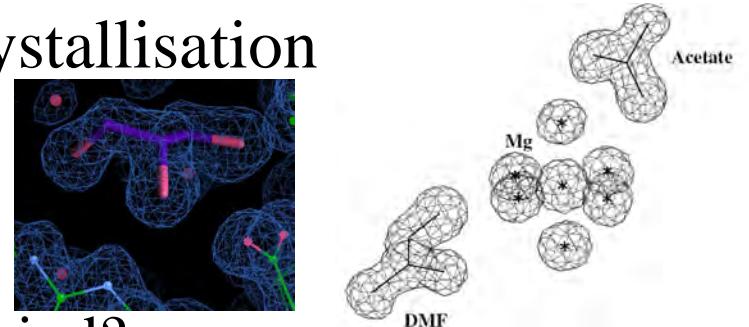
➤ Crystal shape can be misleading

- Bc2lCnt 0.9-1.2 Trisodium citrate pH 7
 - Hexagons OK - flower not OK
- Coda: 1 crystal after several month and broken coverslip
 - Not multiple, 1.3 Å



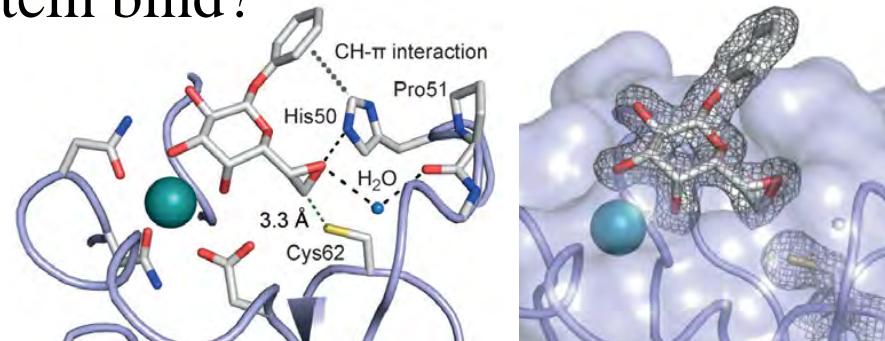
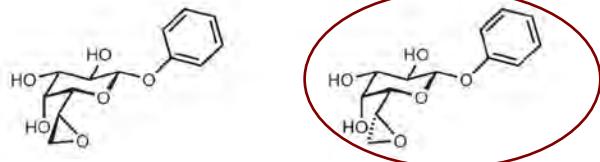
➤ Components from purification or crystallisation

- Electron density of chemical can help
 - Glycerol from cryo in TrfbL1 native
 - Cel6A D405N-SDP5: 1OC7, 1.1 Å



➤ Which diastereoisomer did the protein bind?

- LecA with potential covalent epoxide
 - Crystal at unreactive pH : 4.5



Conclusions



- Still need for X-ray crystallography to gain access to protein-sugar interactions at the atomic level
 - Highest the resolution the better to draw accurate conclusions
 - Distortion / High energy conformation
 - Real or artefact from user/program errors
 - Binding site location and architecture
 - Structure based drug design
 - Site directed mutagenesis: new specificity
 - Quaternary structure and multivalency
 - Gain fundamental knowledge on protein folding and function
 - Design new molecular tools
- Could be tricky for non glycobiologists
 - No « Structural glycobiology for dummies »
 - Do not hesitate to contact experts

Acknowledgements



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Organische Chemie, Zürich

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Alexander Titz
Roman Sommer
Stefanie Wagner
Dirk Hauck

+++

